INTRODUCTION

Approximately 20% of breast cancers (BCs) harbor amplification of the ERBB2 proto-oncogene, yielding overexpression of ERBB2 (HER2) receptor. Before advent of ERBB2-targeted therapies, patients with ERBB2+ tumors experienced poor clinical outcome.1,2 The humanized monoclonal antibody trastuzumab (Herceptin) targets the extracellular domain (ECD) of full-length p185-ERBB2 receptor, and has improved prognosis for many patients with ERBB2+ BC.3–7 However, only subsets of ERBB2+ patients respond to first-line trastuzumab, and resistance to trastuzumab therapy occurs frequently.5,8–10

Several truncated ERBB2 isoforms have been described in human BC, arising via alternative mRNA translation and metalloproteinase cleavage.11,12 Membrane-localized t-ERBB2 isoforms (t-ERBB2s) can activate AKT and mitogen-activated protein kinase signaling in BC cells,13,14 however, they lack the bulk of receptor ECD (including the target epitope of trastuzumab) and may confer trastuzumab resistance.15 indeed, patients with t-ERBB2+ BC exhibit impaired trastuzumab response.13,16 Furthermore, t-ERBB2 expression correlates with increased nodal involvement, and t-ERBB2s are more frequently expressed in metastases than primary tumors.17–19 Importantly, t-ERBB2 expression is associated with shorter progression-free and overall survival of metastatic BC patients, including those treated with trastuzumab.16,18,20

Three t-ERBB2s have been described in clinical specimens and ERBB2-amplified cell lines (Figure 1a): p110 (also referred to as 611CTF17), which arises by alternative translation of ERBB2 mRNA; p95m (m = membrane, also 648CTF), arising via proteolytic cleavage of full-length receptor;21 importantly, both p110 and p95m isoforms contain receptor transmembrane (TM) domain. p95cyto (cytoplasmic, 687CTF), an isoform lacking TM domain, is expressed in the cytoplasm.11,12 Finally, roles for ERBB2 isoforms in cell nuclei have also been described.22–24

Clinical testing for ERBB2 overexpression frequently relies on an intracellular domain-specific antibody (HercepTest; Dako, Carpente-ria, CA, USA),25,26 and thus cannot distinguish between ERBB2 isoforms. If t-ERBB2 expression impacts trastuzumab response,
accurate measurement of t-ERBB2s may influence choice of targeted therapy, for example, therapeutic antibodies vs kinase inhibitors.\textsuperscript{13,16} Therefore, methods distinguishing full-length from t-ERBB2s in a sensitive and specific manner are needed. Herein, we demonstrate a novel antibody-microarray format, Collaborative Enzyme Enhanced Reactive Immunoassay (CEER),\textsuperscript{27} to quantify total and phosphorylated t-ERBB2s in human BC, using frozen primary specimens and fine-needle aspirate (FNA) metastatic biopsies. This method is able to detect and discern full length vs t-ERBB2s with a high degree of sensitivity, using minute amounts of biopsied tumor material.

Having confirmed t-ERBB2 expression in a clinical cohort, we next investigated functional effects of t-ERBB2s in HMLEs.\textsuperscript{28,29} cDNAs encoding ERBB2 isoforms were stably expressed in human mammary epithelial cell lines, HMLE, HME and MCF10A. t-ERBB2 effects on cells were investigated in vitro using soft agar colony formation, and invasion and migration assays. In addition, \textit{in vivo} engraftment of HMLE cells expressing t-ERBB2s was investigated.
using orthotopic xenografts in non-obese diabetic/severe combined immune deficiency mice.

The signal transducer and activator of transcription 5 (STAT5) transcription factor has an important role in directing differentiation of both normal and transformed breast epithelial cells,30 and ERBB receptors (including ERBB2) can regulate phosphorylation of STAT5.31 Levels of phosphorylated STAT5 (pSTAT5) are inversely correlated with BC tumor stage, and pSTAT5 predicts favorable outcome in patients.30,31 In vitro, pSTAT5 inhibits BC cell migration and invasion, potentially explaining its association with favorable prognosis.32,33 Herein, we report attenuation of pSTAT5 accompanying p110-t-ERBB2 expression in multiple BC cell types.

RESULTS

Detection of t-ERBB2s in BC

Full-length and t-ERBB2s are present in cultured BC cell lines SKBr3 and BT474 (Figure 1b). In addition, t-ERBB2s exist in a subset of ERBB2-amplified human BC.34-36 Thus, we sought to confirm the expression of t-ERBB2s in a cohort of BC specimens, using a novel antibody-capture, proximity-based immunomicroarray platform34 (CEER; Figure 1c). This assay requires immunocomplex formation between target-specific capture antibody and two detector antibodies (one conjugated to glucose oxidase and one to horseradish peroxidase (HRP)) by binding to a single target protein.33,35,36 As shown in Figure 1d, this assay detects total and phosphorylated t-ERBB2 using lysates from as few as 250 BT474 cells (see Materials and methods and Supplementary Figure 1A). Comparison of differential ERBB2 profiling (with ERBB2-ECD and ERBB2-intracellular domain (ICD) captures) of BT474 cells with and without the removal of full-length p185-ERBB2 demonstrated that approximately 4.3% of the total ERBB2 receptors were t-ERBB2s (or 5.3 x 10^6 t-ERBB2 receptors per cell) in this ERBB2-amplified cell line, with about 1.2 x 10^7 total ERBB2 receptors per cell. The enrichment of t-ERBB2 was efficient, as removal of p185-ERBB2 from lysates prepared from 250 BT474 cells yielded substantially lower signal with ERBB2-ECD capture antibody, indicating that the signal in ERBB2-ICD antibody capture is predominantly from t-ERBB2. Levels of expression and phosphorylation of t-ERBB2 were determined by comparing values of both capture antibodies on samples before and after p185-ERBB2 clearance by immunoprecipitation. Standard curves of levels of total ERBB2 and phosphorylated ERBB2 were generated as references, using CEER analysis of the BT474 cell line (Supplementary Figure 2).

Frozen BC specimens from 74 patients were scored for ERBB2 levels using standard immunohistochemical analysis (HercepTest) and then analyzed using CEER (Table 1 and Supplementary Table S1). In all, 24 samples were ERBB2 low/negative (score = 0–1 by immunohistochemistry (IHC)), 19 were IHC score 2 and 31 had high-level expression of ERBB2 (IHC score = 3+). The numbers and percentages of samples harboring t-ERBB2 receptors in each IHC group are summarized in Table 1. In this cohort, two IHC –0/1 specimens had detectable full-length ERBB2 above background, but none had t-ERBB2 above background cutoff (p185 level below 300 000 RTK ng^-1) was considered background based on CEER measurement of ERBB2 in T47D non-amplified cell line. In all, 15.7% (3 of 19) of IHC2 ERBB2+ tumors expressed t-ERBB2, whereas 58% (18 of 31) of IHC3+ ERBB2+ tumors expressed t-ERBB2. The association of t-ERBB2 expression with IHC3+ score was statistically significant (IHC3+ cohort compared to others, P<0.001). Furthermore, detection via pan-antiphosphotyrosine antibody revealed that t-ERBB2 isoforms were phosphorylated in subsets of ERBB2 IHC2 tumors (11%, 2 of 19 samples) and IHC3+ tumors (32%, 10 of 31). IHC3+ score was significantly associated with detectable phosphorylated t-ERBB2 (IHC3+ cohort compared to others, P<0.005). CEER-ERBB2 profiling and immunoprecipitation-western blot analyses of ERBB2 isoforms in three individual tumor samples are provided in Figures 2a and b, respectively (values for these samples are highlighted in red; Supplementary Table S1). Supplementary Table S1 provides t-ERBB2 quantification for all 74 samples. Lastly, FNAs of metastatic lesions from eight BC patients were analyzed. Three samples of ERBB2 IHC+ metastases showed varying degrees of total and phosphorylated t-ERBB2 (Table S2), whereas t-ERBB2 were not detected in IHC– metastases (not shown).

Engineered expression of t-ERBB2s

Having confirmed t-ERBB2 expression and phosphorylation in human BC specimens, we sought to determine their biological effects in HMLEs. cDNAs encoding p185, p110, p95m and p95cyto were stably expressed using pLXSN retroviral vector (Figure 3a). In addition, a nuclear isoform (p95n) was constructed by adding two tandem nuclear localization sequences to the p95cyto C terminus (see Figure 1a). Importantly, parental HMLEs lack detectable endogenous t-ERBB2s (Figure 3a). Expression of p185-ERBB2 was used as positive control, while non-transduced and empty-vector-transduced cells served as negative controls in experiments. As shown in Figure 3a, western blotting of HMLE lysates confirmed the expression of ERBB2 isoforms in each stable pool of cells. While protein expression levels varied slightly, comparable levels of p110 compared to p95m and p95cyto compared to p95n were achieved in these pooled populations. HMLE-p110 cells also express the p95cyto isoform due to alternate translation from the downstream AUG codon corresponding to methionine 687 in the ERBB2 peptide sequence (see Figure 3b, upper left panel); however, we have also engineered a version of p110 t-ERBB2 with a mutated 687 methionine codon (M687G) to prevent translation of p95cyto (see western blot; Supplementary Figure 1B). This construct has yielded the same phenotypic effects in cells as the original (data not shown).

Subcellular localization of ERBB2 isoforms

Alternative translation of ERBB2 mRNA is responsible for the generation of p110 (CTF611) t-ERBB2.12 This isoform lacks an N-terminal leader peptide to facilitate canonical membrane protein trafficking.17 We investigated subcellular localization of this isoform using two complementary methods, western blots of cell protein fractions (Figure 3b) and immunofluorescence...
confocal microscopy (Figure 3c). p185, p110 and p95m receptors (those harboring TM domain) were localized to cell membranes. The p95m cDNA used in this study also lacks a canonical N-terminal leader peptide, yet this isoform also resides in the plasma membrane. Thus, it is apparent that the TM domain is sufficient to drive plasma membrane localization of p110 t-ERBB2. In contrast, p95cyto existed predominantly in the cytoplasm, whereas engineered nuclear isoform p95n was predominantly localized to the nucleus (this image is at × 2 higher magnification to enhance detail). Because of the loss of protein during the extraction method for fractionation, some signal is lost for p185 ERBB2 in the p110 and p95m lanes.

p110-t-ERBB2 enhances cell migration and invasion in vitro. Expression of t-ERBB2 has been previously associated with increased nodal involvement and metastasis, suggesting a role for t-ERBB2s in the pathophysiology of BC migration and invasion.

Figure 2. Detection of ERBB2 isoforms in primary human breast tumor samples. (a) CEER analysis of human tumor samples reveals the presence and phosphorylation of t-ERBB2s. Lysates of tumors from three patients with high (no. 26811) or intermediate (nos. 24913 and 25882) ERBB2 expression as scored by IHC were tested. Assay configuration is illustrated below the CEER panels. Antibodies were arrayed in triplicate at three concentrations. Left panel antibodies for total p185 ERBB2 assay: control IgG (pink arrows), ECD-ERBB2 (yellow arrows) and cytokeratin (CK, green arrows). Center and right panel antibodies for total and phosphorylated t-ERBB2 assay: control IgG (pink arrows), ECD-ERBB2 (yellow arrows), ICD-ERBB2 (dark and light blue arrows) and phosphorylated ERBB2 (red arrows). Before t-ERBB2 CEER assay (center and right panels), full-length p185 ERBB2 was removed with an N-terminal ECD-directed antibody. (b) Western blot analysis of ERBB2 isoforms in human breast tumor samples with varying levels of ERBB2 (by IHC) assayed in part (a). Patient specimen lysates 26811, 24913 and 25882 were probed with antibody against ICD-ERBB2. Patient 26811 with positive western analysis also showed high level of ERBB2 as well as t-ERBB2 expression by CEER, with the use of much less cell lysate. Using CEER configuration, a quantitative detection of full-length ERBB2 and t-ERBB2 was achieved in samples (24913 and 25882) with IHC2⁺ status. Of note, t-ERBB2 was not detected by western blot analysis in these samples. Furthermore, a robust phosphorylation of t-ERBB2 was observed in patient 26811, whereas lower level was detected in patient 24913.
Therefore, we analyzed effects of t-ERBB2s using *in vitro* migration and invasion assays. As shown in Figure 4a, p110 expression enhanced migration of HMLEs compared to p185 ERBB2 (p110 compared to p185, *P* = 0.04). While p95m increased migration compared to parental and empty-vector HMLEs, its effect was not significant compared to p185. Neither p95cyto nor p95n significantly affected cell migration. To rule out cell type- or population-specific effects, we also determined effects of p185 and p110 expression in HME and MCF10A cells. p110 t-ERBB2 also significantly increased migration of HME cells (Figure 4c) compared to p185. MCF10A migration was also enhanced by the expression of p110 compared to p185 (mean number of cells per field = 464 (p110) vs 182 (p185), *P* = 0.004 (Student’s t-test)).

Invasion assays using Matrigel-coated filters (Figure 4b) revealed that p110 and p95m isoforms increased HMLE invasion compared to parental and vector control cells (p110 vs controls, *P* = 0.001; p95m vs controls, *P* = 0.003); however, only p110 significantly increased invasion compared to p185-expressing cells (*P* = 0.03). p110-expressing HME cells were also more invasive than p185-expressing cells (Figure 4d). Importantly, the differences seen in these assays were not due to enhanced proliferation by subsets of cells, as there was no significant difference in the proliferation rate of HMLE cells expressing the different isoforms within the 36 h window of this assay (Supplementary Figure 3C).

In addition, the ability of HMLEs expressing different isoforms of ERBB2 to form colonies in soft agar was assayed. The number of colonies formed by HMLEs significantly increased with the expression of membrane-bound isoforms p185 (*P* = 0.0001), p95m (*P* = 0.003) and p110-ERBB2 (*P* = 0.0001) when compared to vector control cells; however, this effect was not elicited by intracellular t-ERBB2 species (Supplementary Figure 3A).

p110 t-ERBB2 promotes HMLE xenograft formation

Transgenic mouse models have implicated p110 (611CTF) in promoting primary breast tumor formation. Thus, we investigated t-ERBB2 effects on orthotopic xenograft formation by HMLE cells. Recombinant HMLE cells were grafted into
mammary fat pads of non-obese diabetic/severe combined immune deficient mice. Cells expressing oncogenic HRAS were used as positive controls for xenograft formation, whereas parental and empty vector cells served as negative controls. As shown in Figure 5a, only HMLEs expressing HRAS or p110-t-ERBB2 formed detectable xenografts within 150 days. HMLE-HRAS xenografts exhibited a shorter latency period (mean time to xenograft detection = 19.5 days; 95% confidence interval: ± 10.0 days) than p110 t-ERBB2 tumors (mean time to detection = 98 days; 95% confidence interval: ± 41.3 days). Images of a p110 HMLE xenograft in situ, ex vivo and a hematoxylin/eosin-stained section are shown in Figure 5b. Polymerase chain reaction using ERBB2-specific forward primer and vector-specific reverse primer demonstrated that genomic DNA from HMLE-p110 tumors was
positive for pLXSN-p110 construct (Figure 5c). Expression of other ERBB2s, including p185, did not lead to xenograft outgrowth within 150 days.

STAT5 phosphorylation is attenuated by p110-t-ERBB2 expression

On the basis of our in vitro and in vivo observations, we sought a biological basis for p110-driven phenotypes. Unexpectedly, we did not observe significant changes in phosphorylation of MEK1/2, ERK1/2 or AKT signaling proteins in p110- compared to p185-expressing HMLE cells when examined by western blotting or CEER (data not shown), in contrast to a published study using engineered expression in MCF7 cells.14 In addition, there was no significant proliferation rate difference between the HMLE populations (Supplementary Figure 3b). To determine potential mechanisms by which p110-t-ERBB2 elicits phenotypes distinct from p185, we used the Proteome Profiler array (R&D Systems, Minneapolis, MN, USA; no. ARY003) to examine phosphorylation of multiple signaling proteins in HMLE cells. Analysis revealed decreased phosphorylation of STAT5 in HMLE cells expressing p110 isoform, compared to p185-expressing and control HMLE cells (Supplementary Figure 3B). Densitometric analysis revealed approximately twofold reduction of pSTAT5 by this method. Moreover, western blotting verified loss of pSTAT5 in p110-expressing HMLE cells (Figure 6a), and CEER measurement confirmed approximate twofold decrease of pSTAT5 in p110-HMLE compared to p185-HMLE (data not shown). pSTAT5 was also decreased in an independent p110-expressing cell line, HME, when compared to expression of p185 ERBB2 (Figure 6b), ruling out population- and cell-type-specific effects. Loss of pSTAT5 has previously been associated with increased tumor grade and poor clinical outcome in human BC30,32 (see Discussion). To confirm that the decrease in pSTAT5 was sufficient to effect STAT5-dependent gene transcription, we analyzed levels of the STAT5-responsive gene p21<sup>WAF1/CIP1</sup>. p21 protein was decreased in p110-expressing HMLE cells compared to p185 and control HMLE cells (Figure 6c).

To determine if decreased pSTAT5 is sufficient to elicit phenotypes similar to those elicited by p110, we transduced...
HMLE cells with short hairpin (shRNA) lentivirus (Open Biosystems, Huntsville, AL, USA) to silence STAT5 expression. As shown in Figure 6d, HMLE cells transduced with shRNA virus have reduced expression of STAT5 protein, compared to control cells transduced with non-silencing control shRNA. Importantly, STAT5-silenced HMLE cells exhibited enhanced migration (Figure 6e, left panels) and invasion (Figure 6e, right panels) when compared to control cells using transwell assays in vitro, mimicking phenotypes observed in p110-t-ERBB2-expressing cells. To confirm the role of pSTAT5 loss in the migration and invasion driven by p110 expression, we expressed a constitutively active mutant form of STAT5b (STAT5b1*6) in HMLE cells also expressing p110 t-ERBB2. Expression of constitutively active STAT5b in p110 HMLE reversed the p110-driven migratory (Figure 7a) and invasive phenotypes (Figure 7b).

**DISCUSSION**

Routine clinical testing of BC does not currently assess the expression of t-ERBB2 vs p185-ERBB2.\(^{37}\) Although t-ERBB2s may confer resistance to trastuzumab therapy, ERBB2 kinase inhibitors including lapatinib inhibit the activity of t-ERBB2s.\(^{38}\) Therefore, sensitive methods to accurately and reliably measure expression of t-ERBB2s in BC are needed, and could potentially impact choice of anti-ERBB2 therapy.

Recently, t-ERBB2 levels in clinical samples have been assessed by antibody-based VeraTag assay.\(^{16}\) This method requires formalin-fixed and paraffin-embedded tissue that may alter antigenicity due to fixation and antigen retrieval procedures. Therefore, detection of minor t-ERBB2 variants against the background of predominant full-length ERBB2 proteins can be challenging.

In contrast, CEER utilizes tissues (FNA or biopsy) without fixation, thus minimizing the probability of antigen modification during processing. Full-length ERBB2 proteins present in sample lysate are removed immunomagnetically using an antibody against the ECD of ERBB2 (as shown in Supplementary Figure 1A), followed by detection of t-ERBB2 variants using three different ICD antibodies. This configuration combined with the superior sensitivity of the proximity-mediated platform allows minute amounts of t-ERBB2 to be efficiently detected and quantified.
when compared to p185.17,39 To rule out cell-type-specific effects, we also confirmed increased migration driven by p110 in MCF10A cells. Finally, only the expression of p110-t-ERBB2 led to HMLE xenograft formation in vivo. Consistent with our data, p110-t-ERBB2 transgenic mice develop mammary adenocarcinomas with a shorter latency than those in p185-ERBB2 transgenic animals.17 Combined, these data indicate that p110 t-ERBB2 is highly tumorigenic compared to p185 and other isoforms,40 identifying p110 as a potential therapeutic target in ERBB2 + BC.

Unexpectedly, expression of t-ERBB2s did not result in hyperactivation of mitogen-activated protein kinase or AKT signaling pathways, as was described in MCF7 cells.14 Importantly, MCF7 are a fully transformed human cell line with an activating mutation in PI3KCA (among other genetic alterations), possibly explaining the differences in our observations using partially transformed HMLE cells. Rather, p110 t-ERBB2 led to decreased pSTAT5 when compared to p185 ERBB2, in both HME and HMLE cells.

The role of STAT5 in human BC is currently controversial and not fully understood. Activated STAT5 may promote survival and antiapoptotic signaling in mammary cells, contributing to initial neoplastic transformation. However, the presence of pSTAT5 is inversely correlated with BC tumor stage.12 STAT5 is involved in differentiation of breast epithelium; thus, tumors with high pSTAT5 tend to be well-differentiated and have more favorable prognostic characteristics compared to those lacking pSTAT5,30 implying a role for STAT5 in the pathophysiology of BC. Accordingly, silencing of STAT5 in HMLE cells led to increased cell migration and invasion, mimicking p110 t-ERBB2 phenotypes, while expression of constitutively active STAT5 reversed p110-driven migration and invasion. Deregulation of STAT5 signaling has been implicated in motility, invasion and metastasis of BC cells.33,41 Taken together, our data offer a mechanism by which p110 t-ERBB2 may augment malignant phenotypes such as migration and invasion, namely via perturbation of STAT5 signaling. Further elucidation of the molecular mechanism by which p110 t-ERBB2 exerts effects on STAT5 phosphorylation is an active area of research in our laboratory.

In summary, these data implicate p110-t-ERBB2 as a potential driver of tumorigenesis, migration and invasion of human mammary epithelial cells (HMECs), thus extending observations from fully transformed BC cell lines and transgenic mice. Perturbation of STAT5 signaling by p110 t-ERBB2 represents a newly identified mechanism of t-ERBB2 activity in human mammary cells. t-ERBB2s may be drivers of ERBB2 + BC pathogenesis, and could serve as biomarkers identifying patients that may differentially benefit from anti-ERBB2 therapies (monoclonal antibodies (Abs) vs TKIs).16,42 As shown here, the specific and sensitive CEER detection method is able to detect and quantify t-ERBB2 isoforms in human breast tumor samples using minute amounts of clinical material, including metastasis samples. Accurate clinical measurement of t-ERBB2 expression in human tumors may help to guide appropriate ERBB2-targeted therapeutic approaches.

**MATERIALS AND METHODS**

**Multiplexed-microarray printing**

Capture Abs were printed on nitrocellulose-coated glass slides (ONCYTE; Grace Bio-Labs, Bend, OR, USA) using non-contact printers (Nanoplotter, GeSiM); spot diameters were approximately 175 μm. Slides were kept in desiccated chambers at 4°C. Spots included tracking dye and specific capture Abs and detector enzymes, glucose oxidase or HRP, and serial dilution concentrations of 1, 0.5 and 0.25 mg ml⁻¹. Purified mouse-IgGs served as negative controls.

**Antibody conjugation**

Target-specific Abs and detector enzymes, glucose oxidase or HRP, were activated with bi-functional crosslinker, succinimidyl-4-(N-
maleimidomethyl cyclohexane-1-carboxylate, and coupled making antibody–enzyme conjugates. Conjugates were high performance liquid chromatography purified. Ab activities in purified conjugates were determined by competition enzyme-linked immunosorbent assay; enzyme activities were detected by functional assays specific for detector enzymes.

**CEER**

Immunoo microarray slides were rinsed twice with TBST (50 mM Tris/150 mM NaCl/0.1% Tween-20, pH 7.2–7.4), blocked with 80 µl Whatman blocking buffer for 1 h at room temperature (RT), and then washed 2 × with TBST. Serially diluted lysate controls in 50 µl dilution buffer (2% bovine serum albumin/0.1% Triton X-100/TBS, pH 7.2–7.4) and samples were added to designated subarrays on slides, and then incubated for 1 h at RT. Slides were washed 4 × for 3 min, detector Abs were added in 80 µl reaction buffer and incubated for 2 h at RT. After TBST wash to remove unbound detector Abs, 80 µl of biotin-tyramide solution (5 µg ml⁻¹ in 50 mM glucose/phosphate-buffered saline (PBS)) prepared from 400 µg ml⁻¹ in ethanol solution (Perkin-Elmer, Waltham, MA, USA) was added and incubated for 15 min in darkness. The covalent coupling of the highly reactive, short-lived biotin-tyramide radicals to nucleophilic residues in the proximity of interaction sites causes local deposition of biotinylated tyramide. Subsequent incubation with fluorescently labeled streptavidin generates signals that can be captured by fluorescence microarray scanner for visualization and quantitation. Glucose-oxidase/HRP-mediated tyramide signal amplification process was terminated by washing with TBST 4 ×, dried and kept in darkness until imaged via microarray scanner.

**t-ERBB2 enrichment**

Full-length p185-ERBB2 receptors were cleared from lysates using magnetic bead-coupled antibodies specific to ECD of ERBB2. Resulting p185-ERBB2 depleted lysates, which contained t-ERBB2 receptor proteins lacking the ECD, were used for subsequent quantification of t-ERBB2 expression and phosphorylation.

**Clinical samples**

Flash-frozen BC tissues were obtained from subjects with invasive ductal carcinoma, stages II or III (ILS Bio, Chestertown, MD, USA). Baseline ERBB2-IHC status was available for all samples. Flash-frozen tissue samples were lysed in 100 µl lysis buffer, kept on ice for 30 min and centrifuged. Protein concentrations of supernatants determined by BCA protein assay kit (Pierce, Rockford, IL, USA) and stored at −80 °C before analysis.

**CEER data analysis**

Slides were scanned at three photomultiplier tubes gain settings to increase effective dynamic range. Background-corrected signal intensities were averaged from spots printed in triplicate. The relative fluorescent value of the respective reagent blank was subtracted from each sample. Several criteria were used to filter data from further analyses, including limits on spot footprints, coefficient of variation for spot replicates, overall pad background and intensity of reagent blank. For each assay, a standard curve was generated from serially diluted BT474 lysates. Data were fit to a five-parameter equation derived as a function of capture–antibody concentration and photomultiplier tubes.

**IP-western blotting**

Cell lysates were incubated with magnetic bead-conjugated antibodies against intracellular domain of ERBB2 overnight at 4 °C. Immunomagnetically enriched lysates were resuspended in sample buffer containing β-mercaptoethanol, boiled 5 min, cooled to RT and loaded onto NuPage (Invitrogen) 4–12% gel. Separated proteins were transferred to nitrocellulose membranes, washed, blocked with 5% milk and incubated with then 2 Abs before detection process using nitro blue tetrazolium chloride/5-bromo-4-chloro-3-indolyl phosphate.

**Cells and culture**

HMLE, HME and MCF10A cells were cultured in mammary epithelial growth medium (Lonza, Walkersville, MD, USA) in 5% CO₂ atmosphere at 37 °C. Cells stably transduced with pLXSN vector encoding recombinant ERBB2 isoforms were selected using 0.5 mg ml⁻¹ (HME and HMLE) or 0.75 mg ml⁻¹ G418 (MCF10A), and were kept as pooled populations. HMLE p110 cells with added pLVX-puro constructs were selected by the addition of 0.5 µg ml⁻¹ puromycin. BT474 and SKBR3 cells were cultured in RPMI media supplemented with 10% fetal bovine serum. For proliferation assays, 2500 HMLE cells were seeded in triplicate wells of 24-well plates. Cells were trypsinized and counted 24 and 72 h later using a Coulter Z1 counter.

**Cloning and vectors**

ERBB2 cDNAs were expressed using pLXSN retroviral vector. p185, p95cyto and p95m were cloned using endogenous start codons from ERBB2 mRNA sequence, whereas an AUG start codon was engineered to initiate translation of p95m. Recombinant pLXSN vectors were packaged using Phoenix cells, virus was harvested, followed by HMLE transduction. Constitutively active STAT5b (STAT5b*16) was cloned and expressed using pLVX-puro vector (Clontech, Mountain View, CA, USA) via similar techniques. GIPZ shRNA lentiviral vectors targeting STAT5 (Open Biosystems). Lentiviral particles were produced by cotransfection of GIPZ vector DNA with packaging plasmids pMD2.G and psPAX2 in 293 T cells. Viruses were harvested and used to transduce HMLE cells using standard techniques.

**Immunofluorescence confocal microscopy**

HMLE were grown on coverslips to ~50% confluence, washed with PBS and fixed in 4% paraformaldehyde for 20 min at RT. Slides were stained using concanavalin-A-Alexa Fluor 488 (Invitrogen; C11252) diluted in PBS for 30 min at RT, followed by PBS wash. After permeabilization with Triton X-100 anti-ERBB2 antibody (3B5; Abcam, Cambridge, MA, USA; AB16901) was applied in 5% bovine serum albumin-PBS for 1 h at RT, followed by anti-mouse Cy5-conjugated antibody (Millipore, Billerica, MA, USA) for 30 min. Coverslips were washed, mounted on microslides and imaged using Zeiss confocal microscope, followed by the Axiovision software processing.

**Subcellular fractionation and western blotting**

HMLE were fractionated using QProteome cell-compartment kit (Qiagen, Valencia, CA, USA; no. 37502) according to the manufacturer’s instructions. SKBR3 and BT474 cells were lysed in RIPA buffer. Lysates were run on 4–12% Bis-Tris gels (Invitrogen) and transferred to nitrocellulose membranes, probed with anti-ERBB2 antibody (AB16901, 3B5; Abcam), anti-NaK-pump antibody (ab7671; Abcam), anti-lamin A/C antibody (sc-7293; Santa Cruz Biotechnology), anti-actin HRP antibody (Abcam) or anti-GAPDH antibody (ab9482; Abcam) for fraction controls. Blots were developed using standard chemiluminescence techniques.

**Soft agar assays**

In all, 5000 HMLEs were seeded in 0.3% Noble agar-media mixture, on a layer of 0.6% agar in six-well plates. Cultures were maintained for 21 days at 37 °C and stained with 0.01% crystal violet in PBS overnight.
staining, numbers of visible colonies were counted. Experiments were performed in triplicate, and one-way analysis of variance was used to determine significance.

Invasion and migration assays

In all, 50,000 serum-starved HMLEs were seeded into upper chambers of transwells (BD Biosciences, Bedford, MA, USA). For invasion assays, filters were overlaid with Matrigel. Media containing 20% fetal bovine serum was added to lower chambers as chemoattractant. After 24 h (migration) or 36 h (invasion), cells on top of filters were scraped away; filters were fixed with methanol and stained with crystal violet. Filters were mounted on microslides, images were taken at ×10 magnification using light microscopy. Migrating/invasive cells were counted for each well. Experiments were performed in triplicate, and one-way analysis of variance and Student’s t-tests were used to determine significance.

In vivo methods

Mammary fat pads 4 and 9 of female non-obese diabetic/severe combined immune deficient mice were injected with 3 × 10^7 HMLE cells: parental group 1, vector control (g2); p185-ERBB2 (g3); p110-t-ERBB2 (g4); p95m-ERBB2 (g5); p95cyto-t-ERBB2 (g6); p95n-t-ERBB2 (g7); and hRAS (g8). Six mice per group were injected, yielding 12 potential xenografts per group. Xenograft formation and growth was monitored until reaching 5% of bodyweight, or for a maximum of 150 days. Cells were injected in 200 μl of 1:1 (v/v) mix of Matrigel/Collagen. Work was performed following Institutional Animal Care and Use Committee guidelines.

Phosphoprotein array

R&D Systems Human phosphokinase array kit no. ARY003, protein lysates from parental HMLE cells and those stably transduced with empty vector, p185ERBB2 or p110-t-ERBB2 were isolated and analyzed according to the manufacturer’s instructions. For densitometric analysis, the ImageJ software (NIH, Bethesda, MD, USA) was used.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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